

Survey for Urine Culture with AST according to CLSI

EQA Provider: ESfEQA GmbH
Heidelberg

Survey Coordinator: Dr. H. Depner

Instructions for Use

Notes:

The Bacteriology scheme is suitable for all medical laboratories that undertake routine bacteriology including the isolation, identification and/or antimicrobial susceptibility testing of bacterial organisms.



The controls are simulated clinical specimens and contain viable bacteria that are categorized as either Risk Group 1 or 2. However, the controls are not of human origin. The controls should only be handled by trained personnel in accordance with local biosafety regulations. After use, all materials should be disposed following biosafety procedures.

Various tasks of the proficiency testing scheme may occasionally be given over to qualified subcontractors. However, ESfEQA is responsible to the participants for the subcontractor's work.

Results from sample analysis may only be disclosed to colleagues from other laboratories after the testing period has been concluded.

By registering for and participating in this EQA participants agree to accept the general terms and conditions of ESfEQA GmbH. These can be retrieved online at www.esfeqa.eu.

1. Intended Use

The samples are intended for use as control material for External Quality Assessment (EQA) in medical laboratories for the following parameters:

- (1) **Sample 1: Identification** of bacterial pathogens. Performance requirements include the re-isolation and identification of significant pathogens in the samples (normally reporting genus and species).

Importantly, dispatched samples for identification may be **pathogen-free**. In such a case, please select/submit "**pathogen-free**" or "**negative**" as result.

Samples can also be mixed cultures containing more than one significant pathogen.

- (2) **Samples 2: Antimicrobial Susceptibility Testing (AST)** of isolates specified in section 3. Performance requirements include the re-isolation of the corresponding organism and antimicrobial susceptibility testing as described in **section 7** and according to current **CLSI** guidelines.

The performance criteria for the AST are met if the participant has both reported a result for at least 5 antimicrobial agents per isolate and achieved at least 80% of the possible score for the reported antimicrobial agents per isolate, the AST results being evaluated according to the scheme below:

Target value	Reported result				
	S	I	SDD	R	unsuitable
S	2	1	1	0	0
I	1	2	1	1	0
SDD	1	1	2	1	0
R	-1	1	1	2	0
unsuitable	-1	-1	-1	2	2

2. Product Description

The controls are either freeze-dried samples of viable microorganisms or liquid samples simulating urine.

Sample 1: BACUC-C_2026_04_a (for Identification only)

Sample 2: BACUC-C_2026_04_b (for AST only)

3. Information on samples for use in AST

Sample 2: *Enterococcus faecium*

4. Storage and Stability

The samples should be stored upright at 2-8 °C. They are stable at least until the deadline for data submission as indicated below. Lyophilized samples should be processed within 10 minutes after reconstitution.

5. Sample Preparation and Analysis

All specimens should be considered infectious and handled appropriately. Aseptic technique and usual precautions for handling the bacterial group studied should be applied throughout sample preparation and analysis. All work including sample preparation should be conducted in a biological safety cabinet.

For additional handling precautions, refer to CLSI documents GP05-A3, GP17-A3, M29-A4, and QMS03-Ed4E (available at www.clsi.org), or the "Technical Rules for Biological Agents - Protective measures for activities involving biological agents in laboratories - TRBA 100" (available at <https://www.baua.de/EN/Home>), or to "Biosafety in Microbiological and Biomedical Laboratories - CDC/NIH - Latest edition", or to the regulations currently in use in your country.

Lyophilized samples: Rehydrate the pellet in 1.5 ml general purpose broth (e.g. trypticase soy broth) according to your lab procedures. Gently agitate the vial occasionally until the lyophilisate is completely rehydrated

and the suspension is homogenous. Afterwards the samples should be treated the same way as patient samples, e.g. used for inoculation of appropriate culture media with a sample from the control according to your lab procedures.

Liquid, ready-to-use samples: Allow the sample to equilibrate to room temperature for 15 minutes prior to testing. Directly before use, agitate the sample by gently inverting the vial several times. The samples should be treated the same way as patient samples and analyzed in accordance with the instructions of the instrument and reagent manufacturer.

6. Identification

Methods that can be used for identification include conventional techniques (e.g. Biochemical Bacteria Identification), mass spectrometry (e.g. MALDI-TOF), nucleic acid testing (NAT) and spectroscopy.

7. Antimicrobial Susceptibility Testing (AST)

Susceptibility testing can be performed according to current **CLSI** guidelines using phenotypic methods that include agar dilution, the antibiotic concentration gradient method (Etest), broth microdilution and the disk diffusion method.

For AST of Sample 2, antibiotics should be selected from the corresponding table displayed below, and AST profiles shall be reported according to current **CLSI** guidelines with test results assigned to one of the following AST categories:

S - susceptible

SDD – susceptible-dose dependent

I - intermediate

R - resistant

In addition, please select “**unsuitable**” for antimicrobial agents that are not appropriate for use against the specific isolate and/or type of infection.

If an antibiotic agent, which is suitable for AST of a particular organism, is not tested by the participant, it shall be indicated as “not measured” (e.g. if the corresponding antibiotic is not available).

Antibiotics for Sample 2
Amoxicillin-clavulanate
Ampicillin
Ciprofloxacin
Daptomycin
Gentamicin (High-Level Screen))
Levofloxacin
Linezolid
Penicillin
Quinupristin-dalfopristin
Rifampin
Streptomycin (High-Level Screen)
Teicoplanin
Trimethoprim-sulfamethoxazole

8. Dates and Submission of Test Results

Testing Period for Samples 1 - 2: 13/10/26 - 03/11/26

Please submit your results electronically to ESfEQA via the web interface <https://teqa-labv2.esfeqa.eu>. Contact your local distributor of ESfEQA programs or ESfEQA

directly if you need assistance with the registration in TEQA. Alternatively, though not preferred, use the fax form provided on the ESfEQA homepage.

When using several methods, instruments or reagents for determining a single parameter (Identification; Antimicrobial Susceptibility Testing), please transmit only the method, instrument and reagent, which ultimately yielded the result for the corresponding parameter.

8.1. For the **Identification** of bacterial isolates, submit one of the following generic methods only:

Biochemical Bacteria Identification, MALDI-TOF, Nucleic Acid Testing, or Spectroscopy.

8.2. For the **AST** of bacterial isolates, submit one of the following generic methods only:

Agar dilution, Antibiotic concentration gradient (Etest), Broth Microdilution or Disk diffusion.

9. Deadline for Data Submission

Deadlines for data submission are (time zone GMT +1):

Samples 1 - 2: **03/11/26**

10. Reports and Certificates

The data will be evaluated by ESfEQA. The individual laboratory reports and certificates can be retrieved online at <https://teqa-labv2.esfeqa.eu>.